Notes and comments

# The $(\alpha/\beta)_8$ -barrel structural domains of $\alpha$ -amylase and old yellow enzyme are discontinued by a similar excursion at the same place of the barrels: Does it mean they are evolutionarily related?

Š. Janeček

Institute of Ecobiology, Slovak Academy of Sciences, SK-81434 Bratislava, Slovakia e-mail: sjanecek @ue.savba.sk

*Key words:*  $(\alpha/\beta)_8$ -barrel enzymes;  $\alpha$ -amylase; old yellow enzyme; structure comparison; evolutionary relatedness.

## Introduction

The family of proteins adopting parallel  $(\alpha/\beta)_8$ -barrel folding motif in their structures has become an attractive subject for evolutionary studies (Farber and Petsko, 1990, Janeček and Baláž, 1993). Almost all of the  $(\alpha/\beta)_8$ -barrels are enzymes involved in various kinds of biochemical reactions. Despite the fact that these proteins contain common  $(\alpha/\beta)_8$ -barrel domain or some disrupted version of it (for a recent review, see Reardon and Farber, 1995), their amino acid sequences, in general, exhibit no detectable sequence similarities. Only a few examples of clear evolutionary relatedness between several pairs of  $(\alpha/\beta)_8$ -barrels have been found, e.g.  $\alpha$ -amylase and cyclodextrin glycosyltransferase (MacGregor and Svensson, 1989) or glycolate oxidase and flavocytochrome  $b_2$  (Lindqvist et al., 1991).

The efforts to join the seemingly unrelated subfamilies of  $(\alpha/\beta)_8$ -barrels sequentially have not led, however, to unambiguous success, mainly when the approaches were based on a production of the amino acid sequence alignments that would be structurally satisfactory. Such alignments of  $(\alpha/\beta)_8$ -barrel sequences (Pickett et al., 1992, Sergeev and Lee, 1994) are perfect from the structural point of view but they do not necessarily result in the homologies. These could be hidden in  $(\alpha/\beta)_8$ -barrels due to their very long evolutionary history. During development the homologous amino acid residues from the primordial barrel(s) might have adopted different functional roles (Janeček and Baláž, 1995). This theory is anchored in a simple idea that a conserved sequence region of an  $(\alpha/\beta)_8$ -barrel enzyme should be more or less conserved also in the equivalent part of structure of the other  $(\alpha/\beta)_8$ -barrel enzymes owing to their mutual evolutionary relatedness (Janeček, 1993). This has recently been illustrated (Janeček and Baláž, 1995) by the catalytic Glu230 of  $\alpha$ -amylase located near the C-terminus of the fifth  $\beta$ -strand of its  $(\alpha/\beta)_8$ -barrel that has functionally related structural equivalents in more than ten other  $(\alpha/\beta)_8$ -barrel enzymes, i.e. the glutamates might have adopted different functional roles.

This study presents an indication of evolutionary relatedness of two different  $(\alpha/\beta)_8$ -barrel subfamilies, starch hydrolases and flavin mononucleotide-dependent enzymes with  $\alpha$ -amylase (EC 3.2.1.1) and old yellow enzyme (EC 1.6.99.1) as representatives, respectively. The  $(\alpha/\beta)_8$ -barrel of  $\alpha$ -amylase (Matsuura et al., 1984, Swift et al., 1991) as well as of all starch hydrolases and related enzymes (for a review, see Janeček, 1994) is interrupted between the third  $\beta$ -strand and the third  $\alpha$ -helix by a long loop that forms, in fact, a small distinct domain. Similar segment of the polypeptide chain protruding out in the structurally equivalent part of the old yellow enzyme's  $(\alpha/\beta)_8$ -barrel (Fox and Karplus, 1994a, 1994b) evoked an idea that these structural excursions from the regular barrel domains could be a joining feature remaining from one of the primordial barrel structures. This view can be supported by the comparison of catalytic amino acid residues of  $\alpha$ -amylase and old yellow enzyme localised also in the structurally equivalent parts of their  $(\alpha/\beta)_8$ -barrels.

# Materials and methods

The information concerning the amino acid sequences and three-dimensional structures of  $\alpha$ -amylase from *Aspergillus oryzae* and old yellow enzyme from *Saccharomyces cerevisiae*, respectively, were retrieved from the SwissProt Sequence Data Base: accession numbers P10529 and Q02899, from the Brookhaven Protein Data Bank: PDB-entries 6TAA and 10YA (kindly provided by Dr. P. A. Karplus, Cornell University, Ithaca) and from the relevant literature (Matsuura et al., 1984, Swift et al., 1991) and (Fox and Karplus, 1994a, 1994b).

The segments of polypeptide chains of  $\alpha$ -amylase and old yellow enzyme forming the excursions protruding out of their  $(\alpha/\beta)_8$ -barrel domains between the third  $\beta$ -strand and the third  $\alpha$ -helix were aligned using the program CLUSTAL (Higgins et al., 1992). The residues forming secondary structural elements were manually tuned, where applicable.

For structural comparison of peptide segments of  $\alpha$ -amylase and old yellow enzyme containing the functionally important amino acid residues, the program HYPERCHEM (Autodesk, Inc.) was used.

#### **Results and discussion**

It has been recently pointed out that the  $(\alpha/\beta)_8$ -barrel protein family may have diverged so far that no detectable sequence homologies appear among its members (Janeček and Baláž, 1995). Therefore, in the search for some evolutionary joining features among different  $(\alpha/\beta)_8$ -barrel enzymes, any trace seeming to be relevant

# 1018

 $(\alpha/\beta)_8$ -barrel structural domains of  $\alpha$ -amylasc

AMY 1	19	<-β-> <α> VAN <b>HMGYD</b> GA <b>GSS</b> VDYSVFKPFSSQDYFHPFFIÇ	<α> <β-> 2NY <b>EDQ</b> TQVEDCW <b>LGDN</b>	
		**	. *	*. *
OYE 1	17	VLGWAAFPDNLARDGLR-YDSASDNVFM	ADAEQEAKAKK	-ANNP <b>QHS</b> LT 162
		<-α> <β>	<>	<β>

Fig. 1. Amino acid sequence alignment of the polypeptide excursions from the  $(\alpha/\beta)_{s}$ -barrels of  $\alpha$ -amylase (AMY) and old yellow enzyme (OYE). Asterisks and dots signify the identical residues and conservative substitutions, respectively. Gaps are indicated by hypens. The secondary structure elements are bolded and indicated above and under the alignment for  $\alpha$ -amylase and old yellow enzyme, respectively.

should not be neglected. One of these evolutionary traces could be just the excursion from the regular  $(\alpha/\beta)_8$ -barrel domains of  $\alpha$ -amylase and old yellow enzyme.

The excursions protrude from their barrels between the third  $\beta$ -strand and the third  $\alpha$ -helix (Matsuura et al., 1984; Fox and Karplus, 1994a) forming a small distinct domain in both cases. A little is known about its function. It determines, for instance, several functional and stability properties that distinguish the barley  $\alpha$ -amylase isozymes (Rodenburg et al., 1994). The amino acid sequence alignment of these segments of the polypeptide chains of  $\alpha$ -amylase and old yellow enzyme is shown in Fig. 1. There are only five identical residues in the alignment but, more importantly, these positions together with the additional 14 conservative substitutions yielded a degree of sequential similarity (the ratio of the sum of identical and similar residues calculated using the number of residues of the smaller enzyme) as high as 41.4%. It is worth mentioning that this value for two sequentially and structurally homologous, clearly related  $(\alpha/\beta)_8$ -barrel enzymes,  $\alpha$ -amylase and cyclodextrin glycosyltransferase, was found to be 34.4% (their alignment is not shown). Moreover, the secondary structural elements from the excursions of  $\alpha$ -amylase and old yellow enzyme mostly match quite well (Fig. 1). And finally, similar excursions are present in the  $(\alpha/\beta)_8$ -barrels of cyclodextrin glycosyltransferase and oligo-1,6-glucosidase that are closely related to  $\alpha$ -amylase (Jespersen et al., 1993) as well as of trimethylamine dehydrogenase that is closely related to old yellow enzyme (Fox and Karplus, 1994b).

To support the possibility presented above regarding the evolutionary relatedness of these two enzymes, their catalytic amino acid residues were investigated. Remarkably, these residues of old yellow enzyme, His191 and Asn194, are located near the C-terminus of the  $\beta$ 4-strand (Fox and Karplus, 1994a) where, in the  $\alpha$ -amylase structure, Asp206 is positioned, one of the three catalytic residues of  $\alpha$ -amylase (Matsuura et al., 1991). This Asp206 is oriented equivalently with the catalytic histidine of old yellow enzyme (Fig. 2). The fact that the backbones of  $\alpha$ -amylase and old yellow enzyme behind the  $\beta$ 4-strand segments do not fit each other, should not be surprising at all since the enzymes are wholly different, one is a starch hydrolase and the other ranks among flavin mononucleotide-dependent enzymes.

The presence of the other functionally important amino acid residues of  $\alpha$ -amylase at or around its strands  $\beta$ 4 should be also taken into account, such as Lys209 and His210 (both at the substrate binding site (Matsuura et al., 1984)) as well as

Janeček

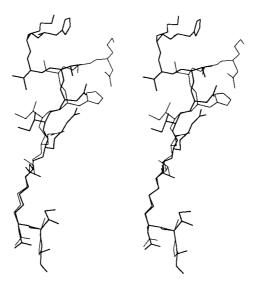


Fig. 2.  $\beta$ 4-Strands of  $\alpha$ -amylase (thick lines) and old yellow enzyme (thin lines) are shown overlapped. Corresponding sequences:  $\alpha$ -amylase: 199\_SIDGLRIDTVKH; old yellow enzyme: 184\_GADGVEIHSANG. Asp206 of  $\alpha$ -amylase and His191 and Asn194 of old yellow enzyme are involved in catalysis of the respective enzymes.

Arg204 (its equivalents in the  $\alpha$ -amylase from pig pancreas and *Bacillus licheni*formis are involved in the chloride binding site (Qian et al., 1993; Machius et al., 1995)). And since the  $\beta$ 4-strand of  $\alpha$ -amylase belongs to one of its highly conserved sequence regions (Janeček et al., 1995), these findings are consistent with the theory on mutual conservation of sequence regions of  $(\alpha/\beta)_8$ -barrel enzymes located in the equivalent parts of their barrel structures (Janeček, 1993).

To summarise, the presented proposal concerning the possible evolutionary relatedness of  $\alpha$ -amylase and old yellow enzyme due to the similarity of their outside  $(\alpha/\beta)_8$ -barrel excursions located in the third loop as well as the similarity between their  $\beta$ 4-strands comprising their catalytic amino acid residues, could make a new connection in the family of all  $(\alpha/\beta)_8$ -barrels. It does not answer the question how these enzymes evolved, but it can support the possibility of a far, divergent evolution road which in the absence of data to the contrary seems more reasonable than a convergence to the sturdy barrel structure (Doolittle, 1995).

## Acknowledgements

The author thanks Dr. P. A. Karplus (Cornell University, Ithaca) for kindly giving the atomic co-ordinates for old yellow enzyme prior their releasing from the Brookhaven Protein Data Bank and Dr. Š. Baláž (Slovak Technical University, Bratislava) for generously providing the facilities necessary for visualising and overlapping protein structures.

1020

 $(\alpha/\beta)_8$ -barrel structural domains of  $\alpha$ -amylase

# References

Doolittle, R. F. 1995. The multiplicity of domains in proteins. Ann. Rev. Biochem. 64: 287-314.

- Farber, G. K. and G. A. Petsko. 1990. The evolution of  $\alpha/\beta$  barrel enzymes. Trends Biochem. Sci. 15: 228–234.
- Fox, K. M. and P. A. Karplus. 1994a. Old yellow enzyme at 2 A resolution: Overall structure, ligand binding, and comparison with related flavoproteins. Structure 2: 1089-1105.
- Fox, K. M. and P. A. Karplus. 1994b. Stirring new interest in an old enzyme: Crystal structure of old yellow enzyme, pp 381–390. In K. Yagi (Ed.), Flavin and Flavoproteins. Walter de Gruyter & Co., Berlin.
- Higgins, D. G., A. J. Bleasby and R. Fuchs. 1992. CLUSTAL V: Improved software for multiple sequence alignment. Comput. Applic. Biosci. 8: 189–191.
- Janeček, Š. 1993. Sequence similarities in  $(\alpha/\beta)_8$ -barrel enzymes revealed by conserved regions of  $\alpha$ -amylase. FEBS Lett. 316: 23-26.
- Janeček, Š. 1994. Parallel  $\beta/\alpha$ -barells of  $\alpha$ -amylase, cyclodextrin glycosyltransferase and oligo-1,6-glucosidase versus the barrel of  $\beta$ -amylase: Evolutionary distance is a reflection of unrelated sequences. FEBS Lett. 353: 119–123.
- Janeček, Š. and Š. Baláž. 1993. Evolution of parallel  $\beta/\alpha$ -barrel enzyme family lightened by structural data on starch-processing enzymes. J. Protein. Chem. 12: 509–514.
- Janeček, Š. and Š. Baláž. 1995. Functionally essential, invariant glutamate near the C-terminus of strand  $\beta$ 5 in various ( $\alpha/\beta$ )<sub>8</sub>-barrel enzymes as a possible indicator of their evolutionary relatedness. Protein Eng. 8: 809–813.
- Janeček, Š., E. A. MacGregor and B. Svensson. 1995. Characteristic differences in the primary structure allow discrimination of cyclodextrin glucanotransferases from α-amylases. Biochem. J. 305: 685–686.
- Lindqvist, Y., C.-I. Brändén, F. S. Mathews and F. Lederer. 1991. Spinach glycolate oxidase and yeast flavocytochrome  $b_2$  are structurally homologous and evolutionarily related enzymes with distinctly different function and flavin mononucleotide binding. J. Biol. Chem. 266: 3198–3207.
- MacGregor, E. A. and B. Svensson. 1989. A super-secondary structure predicted to be common to several  $\alpha$ -1,4-D-glucan-cleaving enzymes. Biochem. J. 259: 145–152.
- Machius, M., G. Wiegand and R. Huber. 1995. Crystal structure of calcium-depleted *Bacillus licheniformis* α-amylase at 2.2 A resolution. J. Mol. Biol. 246: 545-559.
- Matsuura, Y., M. Kusunoki, W. Harada and M. Kakudo. 1984. Structure and possible catalytic residues of Taka-amylase A. J. Biochem. 95: 697–702.
- Matsurra, Y., M. Kusunoki and M. Kakudo. 1991. Structure and catalytic implications of Taka-amylase A. Denpun Kagaku 38: 137-139.
- Pickett, S. D., M. A. S. Saqi and M. J. E. Sternberg. 1992. Evaluation of the sequence template method for protein structure prediction. Discrimination of the  $(\alpha/\beta)_8$ -barrel fold J. Mol. Biol. 228: 170–187.
- Qian, M., R. Haser and F. Payan. 1993. Structure and molecular model refinement of pig pancreatic  $\alpha$ -amylase at 2.1 A resolution. J. Mol. Biol. 231: 785–799.
- Reardon, D. and G. K. Farber. 1995. The structure and evolution of  $\alpha/\beta$  barrel proteins. FASEB J. 9: 497–503.
- Rodenburg, K. W., N. Juge, X.-J. Guo, M. Sogaard, J.-C. Chaix and B. Svensson. 1994. Domain B protruding at the third  $\beta$ -strand of the  $\alpha/\beta$  barrel in barley  $\alpha$ -amylase confers distinct isozyme-specific properties. Eur. J. Biochem. 221: 277–284.
- Sergeev, Y. and B. Lee. 1994. Alignment of beta-barrels in  $(\alpha/\beta)_8$  proteins using hydrogen-bonding pattern. J. Mol. Biol. 244: 168-182.
- Swift, H. J., L. Brady, Z. S. Derewenda, E. J. Dodson, G. G. Dodson, J. P. Turkenburg and A. J. Wilkinson. 1991. Structure and molecular model refinement of *Aspergillus oryzae* (TAKA) α-amylase: An application of the simulated-annealing method. Acta Crystallogr. B47: 535–544.

Received 28 November 1995;

accepted 5 March 1996.

Corresponding Editor: A. Minelli